# Rat GRO/CINC-2α Assay Kit - IBL

96 Well

Please read carefully this instruction prior you use this assay kit.

#### **INSTRUCTIONS FOR USE**

This product is for research use only and is not intended for diagnostic use.

#### **KIT COMPONENT**

| 3 Standard: (Recombinant Rat GRO/CINC-2α) 0.4 4 EIA buffer 36 5 Solution for labeled antibody 17 6 Chromogen: TMB solution 18 7 Stop solution 17   | Well x 1 |
|--|----------|
| 3 Standard: (Recombinant Rat GRO/CINC-2α) 0.4 EIA buffer 3.5 Solution for labeled antibody 1.5 Chromogen: TMB solution 1.5 Stop solution |          |
| 4 EIA buffer 33 5 Solution for labeled antibody 12 6 Chromogen: TMB solution 13 7 Stop solution 13   | .5mL x 1 |
| 5 Solution for labeled antibody 6 Chromogen: TMB solution 7 Stop solution 11   | .5mL x 1 |
| 6 Chromogen: TMB solution 19 Stop solution 19 11   | 30mL x 1 |
| 7 Stop solution 12   | 12mL x 1 |
| •  | 15mL x 1 |
| 8 Wash buffer conc. 5  | 12mL x 1 |
|  | 50mL x 1 |

#### **MEASURING SAMPLES**

Serum and Cell culture supernatant

(Both recombinant and native forms of Rat GRO/CINC-2 $\alpha$  can be detected with the kit.)

#### **PRINCIPLE**

This kit is a solid phase sandwich ELISA (Enzyme-linked Immunosorbent Assay). As a primary antibody is coated on a plate, samples and standard are added into the wells for 1<sup>st</sup> reaction. After the reaction, HRP-conjugated secondary antibody is added into the wells for 2<sup>nd</sup> reaction. After washing away unbound the secondary antibody, Tetra Methyl Benzidine (TMB) is added to the wells and color develops.

### **OPERATING PRECATION**

- 1 Test samples should be measured soon after collection. For storage of samples, store them frozen and do not repeat freeze/thaw cycles. Thaw the test samples at a low temperature and mix them completely before measurement.
- 2 Test samples should be diluted with "4, EIA buffer" contained in this kit.
- 3 Duplicate measurement of test samples and standards is recommended.
- 4 Standard curve should run for each assay.
- 5 Use test samples in neutral pH range. The contaminations of organic solvent may affect the measurement.
- 6 All reagents should be brought to room temperature (R.T.) and mixed completely and gently before use. After mixing them, make sure of no change in quality of the reagents.
- 7 Use only "8, Wash buffer conc." contained in this kit for washing the precoated plate. Insufficient washing may lead to the failure in measurement.
- 8 Fill the wash buffer each well, invert the plate and make sure the liquid is completely removed by shaking it off if you use a washing bottle. Repeat this washing process several times as instructed in order to avoid any insufficient washing process.
- 9 After remove the wash buffer, tapping the plate against a clean paper towel for completely removing the liquid from the wells and make sure the paper towel is not contact with inside of the wells in this process.
- 10 "6, Chromogen TMB solution" should be stored in the dark due to its sensitivity against light. It should be also avoided contact with metals. Required quantity should be prepared into a collecting container for each use.
- After adding TMB solution into the wells, the liquid in the wells gradually changes the color in blue. In this process the plate should be in dark. Remained TMB solution in the collecting container should not be returned into the original bottle of TMB solution to avoid contamination.
- 12 Measurement of O.D. should be done within 30 minutes after addition of "7, Stop solution".
- 13 Storage of HRP conjugated antibody is not recommended. However, if the HRP conjugates do not use at one time, please store it at below -20°C.

# **OPERATION MANUAL AND DOSAGES**

# 1. Materials needed but not supplied.

Plate reader
Test tubes for dilution
Deionized water
Paper towel
Incubator (37°C±1°C)

Micropipette and tip Measuring cylinder and beaker Plate washer Collecting container (i.e. clean disposable test tube) Refrigerator

## 2. Preparation

(1) Preparation of wash buffer

Dilute "8, Wash buffer conc." 40 fold with deionized water. The diluted one is used for the assay as a wash buffer. Adjust the required quantities if

needed.

(2) Preparation of labeled antibody

Add 10.5mL of "5 Solution for labeled antibody" into "2, Labeled antibody" and leave it for 5 minutes and invert and mixed it well for completely dissolving the powder. This operation should be done immediately prior applying the labeled antibody into wells.

(3) Preparation of standard

Add 0.5 mL of deionized water into the vial of "3, Standard" and completely dissolve it. Concentration of the standard is 1200 pg/mL. Prepare 7 test tubes for dilution of the standard and adding 230  $\mu$ L of the EIA buffer into each tube.

Put 230  $\mu$ L of 1200 pg/mL standard into the tube 600 pg/mL (Tube-1) and gently mix it. Afterword, put 230  $\mu$ L of the mixed liquid of tube-1 into the tube 300 pg/mL (Tube-2) and gently mix it. Dilute two fold standard solution in series to set up 7 points of diluted standard between 600 pg/mL and 9.38 pg/mL.

| Tube-1           | 600  | pg/mL |
|------------------|------|-------|
| Tube-2           | 300  | pg/mL |
| Tube-3           | 150  | pg/mL |
| Tube-4           | 75   | pg/mL |
| Tube-4<br>Tube-5 | 37.5 | pg/mL |
| Tube-5<br>Tube-6 |      | . •   |
|                  |      | pg/mL |
| Tube-7           | 9.38 | pg/mL |

(4) Preparation of test samples

Test sample should be diluted with "4, EIA buffer" as the need arises.

#### **3 MEASUREMENT PROCEDURE**

(1) Add test sample blank

Determine wells for test sample blank. Put  $100\mu L$  each of "4, EIA buffer" into the wells.

(2) Add prepared test samples and standard Put 100 μL prepared test samples and 100 μL prepared standard into appropriate wells.

- (3) Incubation with plate lid (1st reaction).
- (4) Washing

Wash the plate with the prepared wash buffer and remove all liquid.

(5) Add prepared labeled antibody

Put 100 µL prepared labeled antibody into the wells.

- (6) Incubation with plate lid (2<sup>nd</sup> reaction).
- (7) Washing

Wash the plate with the prepared wash buffer and remove all liquid completely.

(8) Add "6, Chromogen - TMB solution"

Put 100 µL the TMB solution into the wells.

- (9) Incubation in dark
- (10) Add "7, Stop solution"

Put 100  $\mu L$  the Stop solution into the wells.

(11) Determination of optical density (O.D.)

Remove any dirt or drop of water on the bottom of the plate and confirm there is no bubble on the surface of the liquid. Then, measure the both O.D. of standard and the test samples against a test sample blank.

Measurement wavelength: 450 nm. In case of 2 wavelengths:

Main wavelength is 450nm. Sub-wavelength is between 600 and 650 nm.

Table for measurement procedure

|                          | able for measurement procedure               |                               |                      |  |
|--------------------------|--|-------------------------------|----------------------|--|
|                          | Test samples                                 | Standard                      | Test sample blank    |  |
| Reagents                 | Test samples<br>100 μL                       | Diluted<br>Standard<br>100 μL | EIA buffer<br>100 μL |  |
| 1 <sup>st</sup> reaction | Incubation for                               | r 60 minutes at 37            | °C with plate lid.   |  |
| Washing                  | 4 times (v                                   | vash buffer more t            | han 350 µL)          |  |
| Labeled antibody         | 100 µL                                       | 100 µL                        | 100 µL               |  |
| 2 <sup>nd</sup> reaction | Incubation for 30 minutes at 37              |                               | °C with plate lid.   |  |
| Washing                  | 5 times (v                                   | vash buffer more t            | han 350 µL)          |  |
| TMB solution             | 100 μL                                       | 100 μL                        | 100 µL               |  |
| Chromogenic reaction     | Incubation for 30 minutes at R.T. (shielded) |                               | R.T. (shielded).     |  |
| Stop solution            | 100 μL                                       | 100 μL                        | 100 µL               |  |
| Measuring O.D.           | 450 nm / 600∼650 nm                          |                               |                      |  |

## **CALCULATION OF TEST RESULT**

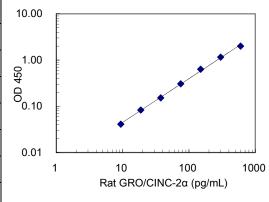
- 1 Plot the concentration of the standard on the x-axis and its O.D. on the y-axis. Draw a standard curve by applying appropriate regression curve on each plot (i.e. quadratic regression of double logarithm conversion).
- 2 Read the concentration by applying the absorbance of the test samples on a standard curve.
- ${\small 3\>\> Calculate\> the\> concentration\> of\> the\> test\> samples\> by\> multiplying\> dilution\> ratio\> of\> test}\\$



samples on the value.

## Example of standard curve and measured value

| Standard<br>(pg/mL) | O.D.<br>(450nm) |
|---------------------|-----------------|
| 600                 | 1.998           |
| 300                 | 1.153           |
| 150                 | 0.632           |
| 75                  | 0.307           |
| 37.5                | 0.152           |
| 18.75               | 0.083           |
| 9.38                | 0.041           |



#### PERFORMANCE AND CHARACTERISTICS

# **Measurement range** 9.38 ∼ 600 pg/mL

2 Dilution linearity

| Test samples    | Dilution<br>ratio<br>(x) | Theoretical<br>value<br>(pg/mL) | Measurement<br>value<br>(pg/mL) | %     |
|-----------------|--------------------------|---------------------------------|---------------------------------|-------|
|                 | 4                        | 292.8                           | 300                             | 97.6  |
| Added 10%FCS    | 8                        | 154.0                           | 150                             | 102.7 |
| Supplemented    | 16                       | 78.0                            | 75                              | 104.0 |
| RPMI-1640 Media | 32                       | 36.5                            | 37.5                            | 97.3  |
|                 | 64                       | 17.0                            | 18.8                            | 90.4  |
|                 | 16                       | 65.1                            | 75                              | 86.8  |
| Serum<br>(Rat)  | 32                       | 37.1                            | 37.5                            | 98.9  |
| (Nat)           | 64                       | 18.1                            | 18.8                            | 96.3  |

3 Added recovery assay

| J | Added recovery assay                   |                                 |                                 |      |
|---|--|---------------------------------|---------------------------------|------|
|   | Test samples                           | Theoretical<br>value<br>(pg/mL) | Measurement<br>value<br>(pg/mL) | %    |
|   |  | 150                             | 138.8 9                         | 92.5 |
|   | Added 10%FCS<br>Supplemented RPMI-1640 | 75                              | 70.8                            | 94.4 |
|   |  | 37.5                            | 34.1                            | 90.9 |
|   |  | 18.8                            | 16.4                            | 87.2 |

4 Intra-assay

| Measurement value (pg/mL) | SD<br>(pg/mL) | CV (%) | n |
|---------------------------|---------------|--------|---|
| 275.4                     | 8.5           | 3.1    | 6 |
| 47.5                      | 2.0           | 4.2    | 6 |
| 16.1                      | 0.8           | 5.0    | 6 |

5 Inter-assay

| o initoi assay            |               |        |   |
|---------------------------|---------------|--------|---|
| Measurement value (pg/mL) | SD<br>(pg/mL) | CV (%) | n |
| 222.2                     | 12.5          | 5.6    | 3 |
| 32.0                      | 4.0           | 12.5   | 3 |

6 Specificity

| Substance       | Cross reactivity (%) |
|-----------------|----------------------|
| Rat GRO/CINC-2α | 100                  |
| Rat GRO/CINC-1  | ≦0.1                 |
| Rat GRO/CINC-2β | ≦0.1                 |
| Rat GRO/CINC-3  | ≦0.1                 |
| Rat MCP-1       | ≦0.1                 |
| Rat Rantes      | ≦0.1                 |
| Rat MIP-1α      | ≦0.1                 |
| Rat IL-1β       | ≦0.1                 |

#### PRECAUTION FOR INTENDED USE AND/OR HANDLING

## 1 Precaution for handling (Hazard prevention)

- (1) Treat the components carefully and wash hands after handling it.
- (2) "7, Stop solution" is a strong acid substance (1N Sulfuric acid). Therefore, it should be careful for the treatment and do not contact your skin and clothes with it. It also needs to pay attention to the disposal of it.

#### 2 Precaution for intended use

- (1) "2, Labeled antibody." And "3, Standard" are lyophilized products. It should be careful to open this vial.
- (2) Should be stored at 2~8°C.
- (3) Precipitation can be seen in "4, EIA buffer" and "8, Wash buffer conc.", however, it does not affect its performance.
- (4) Do not mix or replace the reagents with the reagents from a different lot or kit
- (5) Do not use expired reagents.

#### 3 Precaution for disposal

(1) Dispose used materials after rinsing them with large quantity of water.

#### STORAGE AND THE TERM OF VALIDITY

Storage Condition: 2~8°C

The expiry date is specified on the outer box.

## PACKAGE UNIT AND PRODUCT NUMBER

Package unit: 96 Well Product number: 17170

## **REFERENCES**

1. Makita,H.et al. Effect of anti-macrophage migration inhibitory factor antibody on lipopolysaccharide-induced pulmonary neutrophil accumulation. AMERICAN JOURNAL OF RESPIRATORY AND CRITICAL CARE MEDICINE., 1998: 158(2), 573-579

## **CONTACT DETAILS**

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