

Code No. 27204

**Mouse GIP, Total Assay Kit - IBL****INTRODUCTION**

Incretins are a group of gastrointestinal hormones that cause an increase in the amount of insulin released from the beta cells of the islets of Langerhans after eating and they also inhibit glucagon release from the alpha cells of the Islets of Langerhans. GIP, typical incretin like GLP-1, was isolated and sequenced from intestinal mucosa as "gastric inhibitory peptide" in 1970, and then it was renamed as "glucose-dependent insulintropic peptide". It has been reported that GIP receptor is expressed in cells such as beta cell of pancreas, adipocyte or osteoblastic cell, and it plays essential roles in reserving of ingested nutrients within the body in each cell, and the control of GIP signal can lead to improvement of metabolic syndrome (ref. 1 - 3). GIP is rapidly inactivated to GIP (3-42) from active form of GIP (1-42) by DPP-IV in blood.

This ELISA kit detects both types of mouse GIP, active form (1-42) and inactivated form (3-42), and can measure total GIP in samples.

**PRINCIPLE**

This kit is a solid phase sandwich ELISA using 2 kinds of highly specific antibodies. Tetra Methyl Benzdine (TMB) is used as a coloring agent (Chromogen). The strength of coloring is proportional to the quantities of Mouse total GIP.

**MEASUREMENT RANGE**

12.5 - 800 pmol/L

**INTENDED USE****For research use only, not for use in diagnostic procedures.**

- This IBL's assay kit is capable for the quantitative determination mouse total GIP in EDTA-plasma and cell culture supernatant.

In the case of the active form GIP is also measured with the same sample, DPP-IV inhibitor has to be added when collecting samples, or use purpose-made blood collection tubes in order to preserve GIP. (eg. BD™ P800 Blood Collection System for Preservation of Plasma GLP-1, GIP, Glucagon and Ghrelin by BD).

**KIT COMPONENT**

1	Precoated plate : Anti-GIP (C) Rabbit IgG Affinity Purify	96Well x 1
2	Labeled antibody Conc. : (30X) HRP conjugated Anti- GIP (3-17) (81A1) Mouse IgG MoAb Fab' Affinity Purify	0.4mL x 1
3	Standard : Mouse GIP (1-42) peptide	0.5mL x 2
4	EIA buffer	30mL x 1
5	Solution for Labeled antibody	12mL x 1
6	Chromogen : TMB solution	15mL x 1
7	Stop solution	12mL x 1
8	Wash buffer Conc.	50mL x 1

**OPERATION MANUAL****1. Materials needed but not supplied**

- Plate reader (450nm)
- Graduated cylinder and beaker
- Refrigerator (as 4°C)
- Paper towel
- Incubator (37°C ± 1°C)
- Plate washer or washing bottle\*
- Disposable test tube for "2, Labeled antibody Conc." and "6, Chromogen"
- Micropipette and tip
- Deionized water
- Graph paper (log/log)
- Tube for dilution of Standard

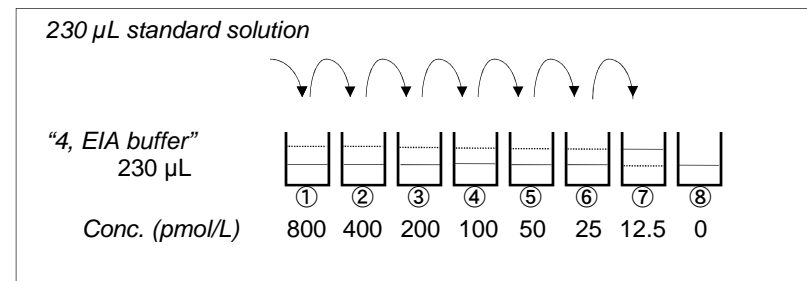
**2. Preparation**

- 1) Preparation of wash buffer  
"8, Wash buffer Conc." is a concentrated (40X) buffer. Adjust the temperature of "8, Washing buffer Conc." to room temperature and then, mix it gently and completely before use. Dilute 50 mL of "8, Wash buffer Conc." with 1,950 mL of deionized water and mix it. This is the wash buffer for use. This prepared wash buffer shall be stored in refrigerator and used within 2 weeks after dilution.
- 2) Preparation of Labeled antibody  
"2, Labeled antibody Conc." is a concentrated (30X). Dilute "2, Labeled antibody Conc." with "5, Solution for Labeled antibody" in 30 times according to required quantity into a disposable test tube. Use this resulting solution as Labeled antibody.  
Example)  
In case you use one strip (8 well), the required quantity of Labeled antibody is 800 µL. (Dilute 30 µL of "2, Labeled antibody Conc." with 870 µL of "5, Solution for Labeled antibody" and mix it. And use the resulting solution by 100 µL in each well.)  
This operation should be done just before applying labeled antibody.  
The remaining "2, Labeled antibody Conc." should be stored at 4°C in firmly sealed vial.
- 3) Preparation of Standard  
Put just 0.5 mL of deionized water into the vial of "3, Standard" and mix it gently and completely. This solution is 1,600 pmol/L Mouse GIP standard.
- 4) Dilution of Standard  
Prepare 8 tubes for dilution of "3, Standard". Put 230 µL each of "4, EIA buffer" into the tube.  
Specify the following concentration of each tube."

Tube-1	800 pmol/L
Tube-2	400 pmol/L
Tube-3	200 pmol/L
Tube-4	100 pmol/L
Tube-5	50 pmol/L
Tube-6	25 pmol/L
Tube-7	12.5 pmol/L
Tube-8	0 pmol/L (Test Sample Blank)

Put 230 µL of Standard solution into tube-1 and mix it gently. Then, put 230 µL of tube-1 mixture into tube-2. Dilute two times standard solution in series to set up 7 points of diluted standard between 800 pmol/L and 12.5 pmol/L. Tube-8 is the test sample blank as 0 pmol/L.

See following picture.

**5) Dilution of test sample**

Test samples have to be diluted with "4, EIA buffer" suitably.

If the concentration of mouse GIP in samples may not be estimated in advance, the pre-assay with several different dilutions will be recommended to determine the proper dilution of samples.

The guide line of dilution rate for plasma sample is 4-fold.

Samples with concentrations higher than that of the highest standard have to be further diluted.

**3. Measurement procedure**

All reagents shall be brought to room temperature approximately 30 minutes before use. Then mix it gently and completely before use. Make sure of no change in quality of the reagents. Standard curve shall be prepared simultaneously with the measurement of test samples.

	Test Sample	Standard	Test Sample Blank	Reagent Blank
Reagents	Test sample 100 µL	Diluted standard (Tube 1-7) 100 µL	EIA buffer (Tube-8) 100 µL	EIA buffer 100 µL
Incubation for 60 minutes at 37°C with plate lid				
4 times (wash buffer more than 350 µL) (Refer to No. 8 and 9 described in OPERATING PRECATION.)*				
Labeled Antibody	100 µL	100 µL	100 µL	-
Incubation for 60 minutes at 4°C with plate lid				
5 times (wash buffer more than 350 µL) (Refer to No. 8 and 9 described in OPERATING PRECATION.)*				
Chromogen	100 µL	100 µL	100 µL	100 µL
Incubation for 30 minutes at room temperature (shielded)				
Stop solution	100 µL	100 µL	100 µL	100 µL
Read the plate at 450nm against a Reagent Blank within 30 minutes after addition of Stop solution.				

- 1) Determine wells for reagent blank. Put 100 µL each of "4, EIA buffer" into the wells.
- 2) Determine wells for test sample blank, test sample and diluted standard. Then, put 100 µL each of test sample blank (tube-8), test sample and dilutions of standard (tube-1-7) into the appropriate wells.
- 3) Incubate the precoated plate for 60 minutes at 37°C after covering it with plate lid.
- 4) Washing (Refer to No. 8 and 9 described in OPERATING PRECATION.)\*  
Wash the plate with the prepared wash buffer and remove all liquid.\*
- 5) Pipette 100 µL of labeled antibody solution into the wells of test samples, diluted standard and test sample blank.
- 6) Incubate the precoated plate for 60 minutes at 4°C after covering it with plate lid.
- 7) Washing (Refer to No. 8 and 9 described in OPERATING PRECATION.)\*  
Wash the plate with the prepared wash buffer and remove all liquid.\*
- 8) Take the required quantity of "6, Chromogen" and put it into a disposable test tube. Then, pipette 100 µL from the test tube into every well. Please do not return the rest of used chromogen in the test tube into "6, Chromogen" bottle in order to avoid contamination.
- 9) Incubate the precoated plate for 30 minutes at room temperature in the dark. The solution of Chromogen will turn blue.
- 10) Add 100 µL of "7, Stop solution" to all wells. Mix the solution by tapping the side of precoated plate. The solution will turn yellow by addition of "7, Stop solution".
- 11) Remove any dirt or drop of water on the bottom of the precoated plate and confirm there is no bubble on the surface of the solution. Then, run the plate reader and conduct measurement at 450 nm against a reagent blank. The measurement shall be done within 30 minutes after addition of "7, Stop solution".

**OPERATING PRECATION**

- 1) Test samples should be measured soon after collection. For storage of samples, store them frozen and do not repeat freeze/thaw cycles. Thaw the test samples at a low temperature and mix them completely before measurement.
- 2) Test samples should be diluted with "4, EIA buffer" contained in this kit.
- 3) Duplicate measurement of test samples and standards is recommended.
- 4) Standard curve should run for each assay.
- 5) Use test samples in neutral pH range. The contaminations of organic solvent may affect the measurement.
- 6) All reagents should be brought to room temperature (R.T.) and mixed completely and gently before use. After mixing them, make sure of no change in quality of the reagents.
- 7) Use only "8, Wash buffer conc." contained in this kit for washing the precoated

plate. Insufficient washing may lead to the failure in measurement.

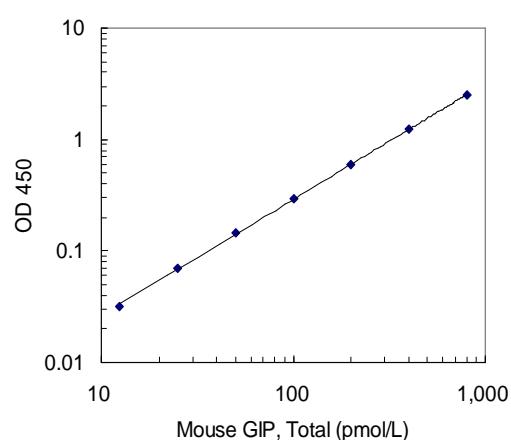
- 8) Using a plate washer is recommended (wait time zero second). It should be washed by a plate washer immediately after each reaction. If you use a washing bottle instead of a plate washer, after filling wash buffer in each well, immediately turn the plate upside down and shake it off to completely remove the wash buffer. Repeat the number of times of wash defined in a table for measurement procedure described in section 3. It should be properly washed off as instructed in order to avoid any insufficient wash.
- 9) Carefully tap the plate against a clean paper towel without contacting with inside of each well to completely remove the washing buffer after repeated the determined number of wash.
- 10) "6, Chromogen - TMB solution" should be stored in the dark due to its sensitivity against light. It should be also avoided contact with metals. Required quantity should be prepared into a collecting container for each use.
- 11) After adding TMB solution into the wells, the liquid in the wells gradually changes the color in blue. In this process the plate should be in dark. Remained TMB solution in the collecting container should not be returned into the original bottle of TMB solution to avoid contamination.
- 12) Measurement of O.D. should be done within 30 minutes after addition of "7, Stop solution".

## CALCULATION OF TEST RESULT

Subtract the absorbance of test sample blank from all data, including standards and unknown samples before plotting. Plot the subtracted absorbance of the standards against the standard concentration on log-log graph paper. Draw the best smooth curve through these points to construct the standard curve. Read the concentration for unknown samples from the standard curve.

Example of standard curve

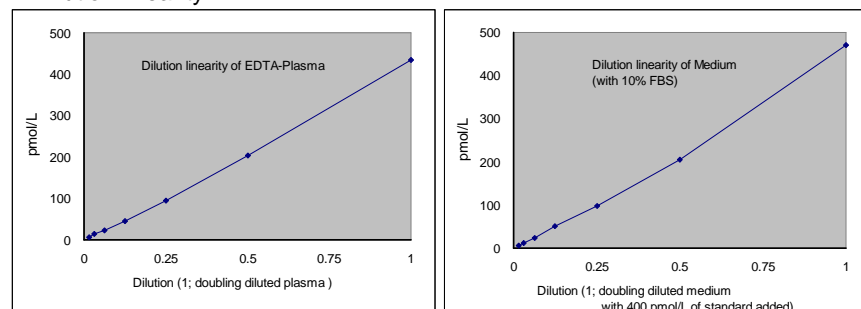
Conc. (pmol/L)	Absorbance (450nm)
800	2.539
400	1.263
200	0.633
100	0.330
50	0.182
25	0.107
12.5	0.069
0 (Test Sample Blank)	0.037



\* The typical standard curve is shown above. This curve can not be used to derive test results. Please run a standard curve for each assay.

## PERFORMANCE CHARACTERISTICS

### 1. Dilution linearity



### 2. Added Recovery Assay

Specimen	Additive Amount (pmol/L)	Theoretical Value (pmol/L)	Measured Value (pmol/L)	%
Mouse Plasma (EDTA) (BALB/c) (x 4)	200	328.3	376.8	114.8
	100	228.3	250.7	109.8
	50	178.3	191.9	107.6
Medium with 10% FBS (x 4)	200	200	201.1	100.6
	50	50	51.1	102.2
	12.5	12.5	13.4	107.2

### 3. Intra - Assay

Mean Value (pmol/L)	SD (pmol/L)	CV (%)	n
324.9	21.4	6.6	24
72.9	5.1	7.0	24
22.7	2.7	11.9	24

### 4. Inter - Assay

Mean Value (pmol/L)	SD (pmol/L)	CV (%)	n
338.2	19.8	5.9	6
77.0	4.7	6.1	6
25.1	2.3	9.2	6

## 5. Specificity

Substance	Cross-Reactivity
Mouse GIP (1-42)	100 %
Mouse GIP (3-42)	100 %
Mouse GLP-1 (7-36) amide	< 0.1 %
Mouse Glucagon	< 0.1 %

## 6. Sensitivity

4.1 pmol/L

The sensitivity for this kit was determined using the guidelines under the National Committee for Clinical Laboratory Standards (NCCLS) Evaluation Protocols. (National Committee for Clinical Laboratory Standards Evaluation Protocols, SC1, (1989) Villanova, PA: NCCLS.)

## PRECAUTION FOR INTENDED USE AND/OR HANDLING

- All reagents should be stored at 2 - 8°C. All reagents shall be brought to room temperature approximately 30 minutes before use.
- "3, Standard" is lyophilized products. Be careful to open this vial.
- "7, Stop solution" is a strong acid substance. Therefore, be careful not to have your skin and clothes contact "7, Stop solution" and pay attention to the disposal of "7, Stop solution".
- Dispose used materials after rinsing them with large quantity of water.
- Precipitation may occur in "2, Labeled antibody Conc.", "4, EIA buffer" or "8, Wash buffer Conc.", however, there is no problem in the performance.
- Wash hands after handling reagents.
- Do not mix the reagents with the reagents from a different lot or kit.
- Do not use expired reagents.
- This kit is for research purpose only. Do not use for clinical diagnosis.

## STORAGE AND THE TERM OF VALIDITY

Storage Condition : 2 - 8°C

The expiry date is specified on outer box.

## REFERENCE

- Miyawaki K, Yamada Y, Yano H, Niwa H, Ban N, Ihara Y, Kubota A, Fujimoto S, Kajikawa M, Kuroe A, Tsuda K, Hashimoto H, Yamashita T, Jomori T, Tashiro F, Miyazaki J, Seino Y. Glucose intolerance caused by a defect in the entero-insular axis: a study in gastric inhibitory polypeptide receptor knockout mice. Proc Natl Acad Sci U S A. 1999 Dec 21;96(26):14843-7.
- Miyawaki K, Yamada Y, Ban N, Ihara Y, Tsukiyama K, Zhou H, Fujimoto S, Oku A, Tsuda K, Toyokuni S, Hiai H, Mizunoya W, Fushiki T, Holst JJ, Makino M, Tashita A, Kobara Y, Tsubamoto Y, Jinnouchi T, Jomori T, Seino Y. Inhibition of gastric inhibitory polypeptide signaling prevents obesity. Nat Med. 2002 Jul;8(7):738-42.
- Tsukiyama K, Yamada Y, Yamada C, Harada N, Kawasaki Y, Ogura M, Bessho K, Li M, Amizuka N, Sato M, Udagawa N, Takahashi N, Tanaka K, Oiso Y, Seino Y. Gastric inhibitory polypeptide as an endogenous factor promoting new bone formation after food ingestion. Mol Endocrinol. 2006 Jul;20(7):1644-51.
- Hansotia T, Maida A, Flock G, Yamada Y, Tsukiyama K, Seino Y, Drucker DJ. Extrapankreatic incretin receptors modulate glucose homeostasis, body weight, and energy expenditure. J Clin Invest. 2007 Jan;117(1):143-52.

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Made in Japan.