

Instruction for Use Code No. 27797

Glucagon ELISA Kit – IBL

96 Well

30mL x 1

50mL x 1

Please read carefully this instruction prior you use this assay kit.

INSTRUCTIONS FOR USE

This product is for research use only and is not intended for diagnostic use.

KIT COMPONENT

| 1 | Precoated plate: (Anti- Glucagon (23-29)* 142A1A Mouse IgG MoAb) | 96Well x 1 |
|---|---|------------|
| 2 | Labeled antibody conc.: | |

- (30X) HRP conjugated Anti- Glucagon N6mer 201A2A Rat IgG Fab') 0.4mL x 1 3 Standard: (Glucagon) 0.5mL x 2
- 4 EIA buffer 5 Solution for labeled antibody
- 5Solution for labeled antibody12mL x 16Chromogen: TMB solution15mL x 17Stop solution12mL x 1
- 7 Stop solution8 Wash buffer conc.

MEASURING SAMPLES

Human EDTA plasma.

% It is recommended to use purpose made blood collection tubes in order to preserve Glucagon.

(eg. BDTM Blood Collection System for Preservation of Plasma GLP-1, GIP, Glucagon and Ghrelin, BD Biosciences)

PRINCIPLE

This kit is a solid phase sandwich ELISA (Enzyme-linked Immunosorbent Assay). As a primary antibody is coated on a plate, samples and standard are added into the wells for 1st reaction. After the reaction, HRP-conjugated secondary antibody is added into the wells for 2nd reaction. After washing away unbound the secondary antibody, Tetra Methyl Benzidine (TMB) is added to the wells and color develops.

OPERATING PRECATION

- 1 Test samples should be measured soon after collection. For storage of samples, store them frozen at -80°C and do not repeat freeze/thaw cycles. Thaw the test samples at a low temperature and mix them completely before measurement.
- 2 Test samples should be diluted with "4, EIA buffer" contained in this kit.
- 3 Duplicate measurement of test samples and standards is recommended.
- 4 Standard curve should run for each assay.
- 5 Use test samples in neutral pH range. The contaminations of organic solvent may affect the measurement.
- 6 All reagents should be brought to room temperature (R.T.) and mixed completely and gently before use. After mixing them, make sure of no change in quality of the reagents.
- 7 Use only "8, Wash buffer conc." contained in this kit for washing the precoated plate. Insufficient washing may lead to the failure in measurement.
- 8 Using a plate washer is recommended (wait time zero second). It should be washed by a plate washer immediately after each reaction. If you use a washing bottle instead of a plate washer, after filling wash buffer in each well, immediately turn the plate upside down and shake it off to completely remove the wash buffer. Repeat the number of times of wash defined in a table for measurement procedure described in section 3. It should be properly washed off as instructed in order to avoid any insufficient wash.
- 9 Carefully tap the plate against a clean paper towel without contacting with inside of each well to completely remove the washing buffer after repeated the determined number of wash.
- 10 "6, Chromogen TMB solution" should be stored in the dark due to its sensitivity against light. It should be also avoided contact with metals. Required quantity should be prepared into a collecting container for each use.
- 11 After adding TMB solution into the wells, the liquid in the wells gradually changes the color in blue. In this process the plate should be in dark. Remained TMB solution in the collecting container should not be returned into the original bottle of TMB solution to avoid contamination.

for the assay as a wash buffer. Adjust the required quantities if needed. (2) Preparation of labeled antibody

Dilute "2, Labeled antibody conc." 30 fold with "5, Solution for labeled antibody" using a prepared collecting container.

Example)

In case you use one strip (8 well), the required quantity of Labeled antibody is 800 μ L. (Dilute 30 μ L of "2, Labeled antibody Conc." with 870 μ L of "5, Solution for labeled antibody" and mix it. And use 100 μ L the mixed solution in each well.) This operation should be done just before applying labeled antibody.

The remaining "2, Labeled antibody Conc." should be stored at 4°C in a firmly sealed vial.

(3) Preparation of standard

Add 0.5 mL of deionized water into the vial of "3, Standard" and completely dissolve it. Concentration of the standard is 40 pmol/L. The standards enclosed in this kit can be frozen and stored after reconstitution. However the freeze-thaw shall not be repeated.

Prepare 7 test tubes for dilution of the standard and adding 230 μL of the EIA buffer into each tube.

Put 230 μ L of 40 pmol/L standard into the tube 20 pmol/L (Tube-1) and gently mix it. Afterword, put 230 μ L of the mixed liquid of tube-1 into the tube 10 pmol/L (Tube-2) and gently mix it. Dilute two fold standard solution in series to set up 7 points of diluted standard between 20 pmol/L and 0.31 pmol/L.

| Tube-1 | 20 | pmol/L |
|--------|------|--------|
| Tube-2 | 10 | pmol/L |
| Tube-3 | 5 | pmol/L |
| Tube-4 | 2.5 | pmol/L |
| Tube-5 | 1.25 | pmol/L |
| Tube-6 | 0.63 | pmol/L |
| Tube-7 | 0.31 | pmol/L |

(4) Preparation of test samples

Dilute test samples with "4, EIA buffer" contained in this kit as follows. Human EDTA plasma : 2 fold

3. Measurement Procedure

(1) Add test sample blank

Determine wells for test sample blank. Put 100µL each of "4, EIA buffer" into the wells.

(2) Add prepared test samples and standard

Put 100 μ L prepared test samples and 100 μ L prepared standard into appropriate wells.

- (3) Incubation with plate lid (1st reaction).
- (4) Washing (Refer to No. 8 and 9 described in OPERATING PRECATION.) Wash the plate with the prepared wash buffer and remove all liquid.
- (5) Add prepared labeled antibody
 - Put 100 µL prepared labeled antibody into the wells.
- (6) Incubation with plate lid (2nd reaction).
- (7) Washing (Refer to No. 8 and 9 described in OPERATING PRECATION.) Wash the plate with the prepared wash buffer and remove all liquid completely.
- (8) Add "6, Chromogen TMB solution"
 - Put 100 μ L the TMB solution into the wells.
- (9) Incubation in dark
- (10) Add "7, Stop solution"

Put 100 μ L the Stop solution into the wells.

(11) Determination of optical density (O.D.)

Remove any dirt or drop of water on the bottom of the plate and confirm there is no bubble on the surface of the liquid. Then, measure the both O.D. of standard and the test samples against a test sample blank.

Measurement wavelength: 450 nm. In case of 2 wavelengths:

Main wavelength is 450nm. Sub-wavelength is between 600 and 650 nm.

| Table for measurement procedure | | | | |
|---------------------------------|---|-------------------------------|----------------------|--|
| | Test samples | Standard | Test sample blank | |
| Reagents | Test samples 100 μL | Diluted Standard 100 µL | EIA buffer 100 μL | |
| 1st reaction | Incubation for a | overnight at 2~8°C | with plate lid | |
| Washing | 2 times (wash buffer more than 350 μL) (Refer to No. 8 and 9 described in OPERATING PRECATION.) | | | |
| Labeled antibody | 100 μL 100 μL 100 μL | | | |
| 2nd reaction | Incubation for 60 minutes at 2~8°C with plate lid | | | |
| Washing | 3 times (wash buffer more than 350 μL) (Refer to No. 8 and 9 described in OPERATING PRECATION.) | | | |
| TMB solution | 100 µL | 100 µL | 100 µL | |
| Chromogenic reaction | Incubation for 30 minutes at R.T. (shielded). | | | |

- 12 Measurement of O.D. should be done within 30 minutes after addition of "7, Stop solution".

OPERATION MANUAL AND DOSAGES

1. Materials needed but not supplied.

| Plate reader | Micropipette and tip |
|-------------------------|-----------------------------------|
| Test tubes for dilution | Measuring cylinder and beaker |
| Deionized water | Plate washer or washing bottle |
| Paper towel | Collecting container |
| Refrigerator | (i.e. clean disposable test tube) |
| | |

2. Preparation

(1) Preparation of wash buffer

Dilute "8, Wash buffer conc." 40 fold with deionized water. The diluted one is used

Manufacturer: Immuno-Biological Laboratories Co., Ltd.

Instruction for Use Code No. 27797

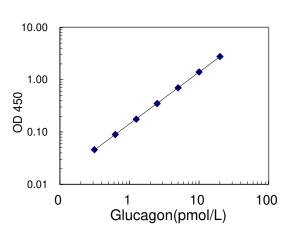
| Stop solution | 100 µL | 100 µL | 100 µL |
|----------------|---------------------|--------|--------|
| Measuring O.D. | 450 nm / 600~650 nm | | nm |

CALCULATION OF TEST RESULT

- 1 Plot the concentration of the standard on the x-axis and its O.D. on the y-axis. Draw a standard curve by applying appropriate regression curve on each plot (i.e. quadratic regression of double logarithm conversion).
- 2 Read the concentration by applying the absorbance of the test samples on a standard curve.
- 3 Calculate the concentration of the test samples by multiplying dilution ratio of test samples on the value.

Example of standard curve and measured value

| Standard (pmol/L) | O.D. (450nm) | |
|-----------------------|-----------------|--|
| 20. | 2.763 | |
| 10. | 1.402 | |
| 5. | 0.698 | |
| 2.5 | 0.350 | |
| 1.25 | 0.176 | |
| 0.63 | 0.090 | |
| 0.31 | 0.046 | |



PERFORMANCE AND CHARACTERISTICS

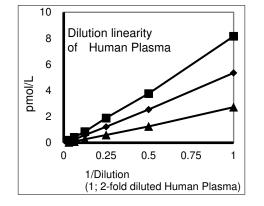
1 Sensitivity

0.05 pmol/L (Calculated by NCCLS method using the standard.)

2 Measurement range

0.31 ~ 20 pmol/L

3 Dilution linearity



4 Added recovery assay

| Specimen | Additive Amount (pmol/L) | Theoretical Value (pmol/L) | Measurement Value (pmol/L) | % |
|------------|--------------------------------|----------------------------------|----------------------------------|-------|
| Human | 10.00 | 11.95 | 12.52 | 104.8 |
| EDTA- | 2.50 | 4.45 | 4.41 | 99.1 |
| Plasma(x2) | 0.63 | 2.58 | 2.40 | 93.0 |

7 Interfering Substances

Hemolyzed hemoglobin does not affect on the value of measurement up to 54 $\rm mg/dL.$

Free bilirubin does not affect on the value of measurement up to 20 mg/dL. Conjugated bilirubin does not affect on the valueof measurement up to 20 mg/dL. Chyle does not affect on the value of measurement up to 1,410 FTU.

8 Specificity

| Substance | Cross reactivity (%) |
|------------------|----------------------|
| Oxyntomodulin | 0.06 |
| Glicentin(1-61) | 0.05 |
| Glucagon(3-29) | <0.01 |
| Glicentin(1-69) | <0.01 |
| Mini-glucagon | <0.01 |
| GLP-1(7-36)amide | 0.01 |
| GLP-1(9-36)amide | 0.01 |
| GLP-2 | <0.01 |
| GIP(1-42) | <0.01 |
| GIP(3-42) | <0.01 |

9 Calibration Reference Material

WHO International Standard Glucagon, Porcine NIBSC code;69/194

PRECAUTION FOR INTENDED USE AND/OR HANDLING

1 Precaution for handling (Hazard prevention)

- (1) Treat the components carefully and wash hands after handling it.
- (2) "7, Stop solution" is a strong acid substance (1N Sulfuric acid). Therefore, it should be careful for the treatment and do not contact your skin and clothes with it. It also needs to pay attention to the disposal of it.

2 Precaution for intended use

(1) "3, Standard" is lyophilized products. It should be careful to open this vial.

- (2) All reagents should be stored at 2 8°C.
- (3) Precipitation can be seen in "4, EIA buffer", "5, Solution for labeled antibody" and "8, Wash buffer conc.", however, it does not affect its performance.
- (4) Do not mix or replace the reagents with the reagents from a different lot or kit.
- (5) Do not use expired reagents.

3 Precaution for disposal

(1) Dispose used materials after rinsing them with large quantity of water.

REFERENCE

1. Kobayashi M, Maruyama N, Yamamoto Y, Togawa T, Ida T, Yoshida M, Miyazato M, Kitada M, Hayashi Y, Kashiwagi A, Kitamura T.A newly developed glucagon sandwich ELISA is useful for more accurate glucagon evaluation than the currently used sandwich ELISA in subjects with elevated plasma proglucagon-derived peptide levels.

J Diabetes Investig. 2023 May;14(5);648-658.

STORAGE AND THE TERM OF VALIDITY

Storage Condition: 2 - 8°C The expiry date is specified on the outer box.

PACKAGE UNIT AND PRODUCT NUMBER

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5 Intra-assay

| Measurement value (pmol/L) | SD(pmol/L) | CV (%) | n |
|-------------------------------|------------|--------|----|
| 11.00 | 0.43 | 3.9 | 24 |
| 2.73 | 0.10 | 3.7 | 24 |
| 0.73 | 0.05 | 6.8 | 24 |

6 Inter-assay

| U Intel abouy | | | |
|----------------------------|-------------|--------|---|
| Measurement value (pmol/L) | SD (pmol/L) | CV (%) | n |
| 11.17 | 0.23 | 2.1 | 8 |
| 2.84 | 0.09 | 3.2 | 8 |
| 0.75 | 0.04 | 5.3 | 8 |

Manufacturer: Immuno-Biological Laboratories Co., Ltd.

Package unit: 96 Well Product number: 27797

CONTACT DETAILS

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