Instruction for Use Code No. 27803

# K-297 Δcbh1 HCP ELISA Kit - IBL

96 Well

Please read carefully this instruction prior you use this assay kit.

### **INSTRUCTIONS FOR USE**

This product is for research use only and is not intended for diagnostic use.

### KIT COMPONENT

1	Precoated plate: (Anti-K-297 Δcbh1 HCP Rabbit IgG)	96Well x 1
2	Labeled antibody conc.:	
	(30X) HRP conjugated Anti-K-297 Δcbh1 HCP Rabbit IgG Fab')	0.4mL x 1
3	Standard: (K-297 \( \Delta cbh1 \) HCP)	0.5mL x 2
4	EIA buffer	30mL x 1
5	Solution for labeled antibody	12mL x 1
6	Chromogen: TMB solution	15mL x 1
7	Stop solution	12mL x 1
8	Wash buffer conc.	50mL x 1

### **MEASURING SAMPLES**

This kit can measure HCP (Host Cell Protein) derived from cell culture medium in TALAMAX® System\*.It can detect HCPs in culture supernatants, intermediate purified products, and final purified products.This kit was developed using culture supernatant-derived HCP that does not contain Cbh1 protein as the immunogen.

\*TALAMAX® System: This is a novel protein secretion expression system developed using the filamentous fungus Talaromyces cellulolyticus as a host for antibody and biopharmaceutical protein production.

### **PRINCIPLE**

This kit is a solid phase sandwich ELISA (Enzyme-linked Immunosorbent Assay). As a primary antibody is coated on a plate, samples and standard are added into the wells for 1<sup>st</sup> reaction. After the reaction, HRP-conjugated secondary antibody is added into the wells for 2<sup>nd</sup> reaction. After washing away unbound the secondary antibody, Tetra Methyl Benzidine (TMB) is added to the wells and color develops.

## **OPERATING PRECATION**

- 1 Test samples should be measured soon after collection. For storage of samples, store them frozen and do not repeat freeze/thaw cycles. Thaw the test samples at a low temperature and mix them completely before measurement.
- 2 Test samples should be diluted with "4, EIA buffer" contained in this kit.
- 3 Duplicate measurement of test samples and standards is recommended.
- 4 Standard curve should run for each assay.
- 5 Use test samples in neutral pH range. The contaminations of organic solvent may affect the measurement.
- 6 All reagents should be brought to room temperature (R.T.) and mixed completely and gently before use. After mixing them, make sure of no change in quality of the reagents.
- 7 Use only "8, Wash buffer conc." contained in this kit for washing the precoated plate. Insufficient washing may lead to the failure in measurement.
- 8 Using a plate washer is recommended (wait time zero second). It should be washed by a plate washer immediately after each reaction. If you use a washing bottle instead of a plate washer, after filling wash buffer in each well, immediately turn the plate upside down and shake it off to completely remove the wash buffer. Repeat the number of times of wash defined in a table for measurement procedure described in section 3. It should be properly washed off as instructed in order to avoid any insufficient wash.
- 9 Carefully tap the plate against a clean paper towel without contacting with inside of each well to completely remove the washing buffer after repeated the determined number of wash.
- 10 "6, Chromogen TMB solution" should be stored in the dark due to its sensitivity against light. It should be also avoided contact with metals. Required quantity should be prepared into a collecting container for each use.
- After adding TMB solution into the wells, the liquid in the wells gradually changes the color in blue. In this process the plate should be in dark. Remained TMB solution in the collecting container should not be returned into the original bottle of TMB solution to avoid contamination.
- 12 Measurement of O.D. should be done within 30 minutes after addition of "7, Stop solution".

# **OPERATION MANUAL AND DOSAGES**

# 1. Materials needed but not supplied.

Plate reader
Test tubes for dilution
Deionized water
Paper towel
Refrigerator
Incubator(37°C±1°C)

Micropipette and tip Measuring cylinder and beaker Plate washer or washing bottle Collecting container (i.e. clean disposable test tube)

# 2. Preparation

(1) Preparation of wash buffer

Dilute "8, Wash buffer conc." 40 fold with deionized water. The diluted one is used for the assay as a wash buffer. Adjust the required quantities if needed.

### (2) Preparation of labeled antibody

Dilute "2, Labeled antibody conc." 30 fold with "5, Solution for labeled antibody" using a prepared collecting container.

### Example)

In case you use one strip (8 well), the required quantity of Labeled antibody is 800  $\mu L$ . (Dilute 30  $\mu L$  of "2, Labeled antibody Conc." with 870  $\mu L$  of "5, Solution for labeled antibody" and mix it. And use  $100\mu L$  the mixed solution in each well.) This operation should be done just before applying labeled antibody.

The remaining "2, Labeled antibody Conc." should be stored at 2~8°C in a firmly sealed vial.

#### (3) Preparation of standard

Add 0.5 mL of deionized water into the vial of "3, Standard" and completely dissolve it. Concentration of the standard is 20 ng/mL. The standards enclosed in this kit can be frozen and stored after reconstitution. However the freeze-thaw shall not be repeated.

Prepare 7 test tubes for dilution of the standard and adding 230  $\mu$ L of the EIA buffer into each tube.

Put 230  $\mu$ L of 20 ng/mL standard into the tube 10 ng/mL (Tube-1) and gently mix it. Afterword, put 230  $\mu$ L of the mixed liquid of tube-1 into the tube 5 ng/mL (Tube-2) and gently mix it. Dilute two fold standard solution in series to set up 7 points of diluted standard between 10 ng/mL and 0.16 ng/mL.

Tube-1	10	ng/mL
Tube-2	5	ng/mL
Tube-3	2.5	ng/mL
Tube-4	1.25	ng/mL
Tube-5	0.63	ng/mL
Tube-6	0.31	ng/mL
Tube-7	0.16	ng/mL

## (4) Preparation of test samples

Dilute test samples with "4, EIA buffer" contained in this kit as follows.

### 3. Measurement Procedure

(1) Add test sample blank

Determine wells for test sample blank. Put 100µL each of "4, EIA buffer" into the wells.

(2) Add prepared test samples and standard

Put 100  $\mu L$  prepared test samples and 100  $\mu L$  prepared standard into appropriate wells

- (3) Incubation with plate lid (1st reaction).
- (4) Washing (Refer to No. 8 and 9 described in OPERATING PRECATION.) Wash the plate with the prepared wash buffer and remove all liquid.
- (5) Add prepared labeled antibody

Put 100 µL prepared labeled antibody into the wells.

- (6) Incubation with plate lid (2nd reaction).
- (7) Washing (Refer to No. 8 and 9 described in OPERATING PRECATION.)

  Wash the plate with the prepared wash buffer and remove all liquid completely.
- (8) Add "6, Chromogen TMB solution"

Put 100  $\mu L$  the TMB solution into the wells.

- (9) Incubation in dark
- (10) Add "7, Stop solution"

Put 100 μL the Stop solution into the wells.

(11) Determination of optical density (O.D.)

Remove any dirt or drop of water on the bottom of the plate and confirm there is no bubble on the surface of the liquid. Then, measure the both O.D. of standard and the test samples against a test sample blank.

Measurement wavelength: 450 nm. In case of 2 wavelengths:

Main wavelength is 450nm. Sub-wavelength is between 600 and 650 nm.

Table for measurement procedure

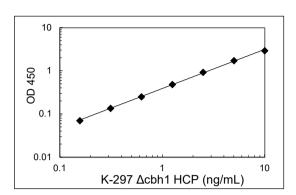
l able for measurement procedure			
	Test samples	Standard	Test sample blank
Reagents	Test samples 100 μL	Diluted Standard 100 μL	EIA buffer 100 μL
1st reaction	Incubation for 6	60 minutes at 37°0	with plate lid
Washing	(Refer to	ash buffer more th o No. 8 and 9 des RATING PRECAT	cribed in
Labeled antibody	100 µL	100 μL	100 µL
2nd reaction	Incubation for 30 minutes at 37°C with plate li		C with plate lid.
Washing	5 times (wash buffer more than 350 μL) (Refer to No. 8 and 9 described in OPERATING PRECATION.)		
TMB solution	100 μL	100 μL	100 μL
Chromogenic reaction	Incubation fo	r 30 minutes at R.	T. (shielded).
Stop solution	100 μL	100 μL	100 μL
Measuring O.D.	450 nm / 600~650 nm		

## **CALCULATION OF TEST RESULT**

- 1 Plot the concentration of the standard on the x-axis and its O.D. on the y-axis. Draw a standard curve by applying appropriate regression curve on each plot (i.e. quadratic regression of double logarithm conversion).
- 2 Read the concentration by applying the absorbance of the test samples on a standard curve.
- 3 Calculate the concentration of the test samples by multiplying dilution ratio of test samples on the value.

Example of standard curve and measured value

Standard ( ng/mL)	O.D. (450nm)
10	2.924
5	1.734
2.5	0.928
1.25	0.485
0.63	0.251
0.31	0.137
0.16	0.070



### PERFORMANCE AND CHARACTERISTICS

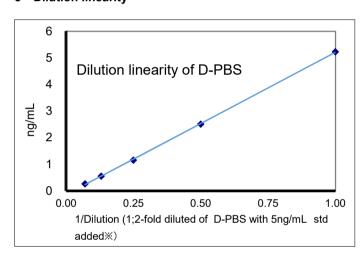
### 1 Sensitivity

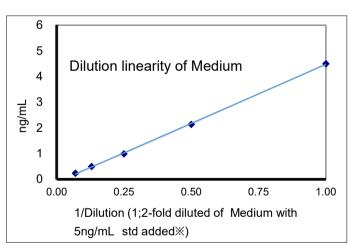
0.02 ng/mL (Calculated by NCCLS method using the standard.)

## 2 Measurement range

0.16  $\sim$  10 ng/mL

## 3 Dilution linearity





(%Samples with standard added are used.)

4 Added recovery assay

Specimen	Additive Amount (ng/mL)	Theoretical Value (ng/mL)	Measurement Value (ng/mL)	%
	5.00	5.00	5.34	106.8
D-PBS(x2)	2.50	2.50	2.65	106.0
D-PB3(XZ)	1.25	1.25	1.25	100.0
	0.63	0.63	0.57	90.5
	5.00	5.00	4.91	98.2
Medium	2.50	2.50	2.20	88.0
(x2)	1.25	1.25	0.99	79.2
	0.63	0.63	0.49	77.8

5 Intra-assay

Measurement value (ng/mL)	SD(ng/mL)	CV (%)	n
4.30	0.13	3.0	24
2.09	0.09	4.3	24
0.76	0.04	5.3	24

6 Inter-assay

o ilitor-accay			
Measurement value (ng/mL)	SD (ng/mL)	CV (%)	n
4.10	0.30	7.3	7
1.97	0.25	12.7	7
0.74	0.06	8.1	7

7 Specificity

r opecinicity		
Substance	Cross reactivity (%)	
K-297 Δcbh1 HCP	100%	
D-PBS	N.D.	
Medium	N.D.	
Highly concentrated protein solution	N.D.	

### PRECAUTION FOR INTENDED USE AND/OR HANDLING

## 1 Precaution for handling (Hazard prevention)

- (1) Treat the components carefully and wash hands after handling it.
- (2) "7, Stop solution" is a strong acid substance (1N Sulfuric acid). Therefore, it should be careful for the treatment and do not contact your skin and clothes with it. It also needs to pay attention to the disposal of it.

#### 2 Precaution for intended use

- (1) "3, Standard" is lyophilized products. It should be careful to open this vial.
- (2) All reagents should be stored at 2 8°C.
- (3) Precipitation can be seen in "4, EIA buffer", "5, Solution for labeled antibody" and "8, Wash buffer conc.", however, it does not affect its performance.
- (4) Do not mix or replace the reagents with the reagents from a different lot or kit.
- (5) Do not use expired reagents.

# 3 Precaution for disposal

(1) Dispose used materials after rinsing them with large quantity of water.

# STORAGE AND THE TERM OF VALIDITY

Storage Condition: 2 - 8°C

The expiry date is specified on the outer box.

# PACKAGE UNIT AND PRODUCT NUMBER

Package unit: 96 Well Product number: 27803

# REFERENCE

# CONTACT DETAILS

Immuno-Biological Laboratories Co., Ltd.

1091-1 Naka, Fujioka-Shi, Gunma 375-0005, Japan
URL: https://www.ibl-japan.co.jp/en/ E-mail: do-ibl@ibl-japan.co.jp